lay it down on the mud surface in a way not to include air bells, The cloth will in a few moments become wet and may then be raised by one corner first and folded up with the side that was next to the mud on the inside. After folding wrap in waxed paper and label for future reference. When ready to clean, place the cloth in a porcelain evaporating dish and cover with strong sulphuric acid and enough bi-chromate of soda to make the mass a deep reddish color. Place the dish in a sand bath over a gas stove or other source of heat, boil the mass till crystal of chromic acid appear as a scum on the surface of liquid. Remove and let cool and pour into a preserve jar partly filled with water. Let settle for at least one-half hour undisturbed, then siphon off water with a rubber tube to within one inch of the bottom of the jar, being careful not to disturb the sediment. Repeat the washing till clear from all color. The sediment may now be removed to a small bottle and examined and if a small quantity of sand is present it may be removed by whirling it with some water in the evaporating dish by means of a glass rod, and the sand will be found to pile up in the center as a dark spot. Carefully pour off the water with the diatoms suspended in it, leaving the sand in the dish.

One will be surprised how the diatoms will stick to the cloth and how little foreign matter will be collected by this method. The above method may be used in fresh water streams provided the spot is first drained.

Providence, R. I. OLIVER KENDALL, JR.

NEW METHOD OF EXAMINING STOOLS FOR EGGS.

C. M. Fauntleroy and R. Hayden (U. S. Naval Med. Bul. Jan. 1915) suggest the following method:

1. Mix thoroughly about 2 grams of fæcal matter with 5 cc. of a 2% aqueous solution of lysol in a centrifuge tube.

2. Centrifugalize at high speed for one minute, decant the supernatant liquid, and mix a fresh quantity of the lysol solution with the sediment in the tubes. Repeat this step three times.

3. Remove small portions of centrifugalized sediment with pipette; place on slide; mix a small drop of anilin gentian violet with the sediment; cover and examine.

The authors have used the method successfully in more than a thousand cases. All eggs, hookworms and others, stand out very clearly. Everything is stained except the eggs. These appear in the natural state because their membranes resist the stain, as they would not in an alcoholic solution, which enables one to run a slide through quickly with the certainty that no eggs are overlooked.

The lysol is not essential but is used as a precaution for disinfection. The method facilitates this disagreeable work, and is so sure that much of it can be turned over to less expert assistants. The clearness of differentiation saves much time.

Abstracted by V. A. LATHAM.

A CLEARING FLUID FOR CELLOIDIN.

Dissolve 1 oz. absolute phenol-crystals in a mixture of equal parts of oils of cedar and bergamot, using gentle heat to hasten the solution. Sections may be cleared in this direct from ordinary methylated spirit. It can be used several times.

V. A. L.

BOUIN'S FLUID: A GOOD ALL-ROUND FIXING SOLUTION

Corrosive sublimate	40 grains
Water	4 ounces
Picric acid	60 grains
Formalin	8 ounces

Dissolve corrosive sublimate in hot water. When cold add picric acid, and then the formalin. Wash with water.

V. A. L.

MOUNTING ZOOPHYTES AND POLYZOA.

Fix and stain in any suitable way. Select suitable branches, place in a little vessel of oil of cloves or other clearing agent, but never let dry. Mount in sunken cell-slip without pressure or distortion. To do so *overfill* the cell, slowly slide the cover over the fluid, and put a clip on each edge of slide. If the clip bears on the center it may crack the cover, or when it is removed the cover may reassert its level and draw in a bubble of air. Larval stages of